

RESEARCH ARTICLE

Open Access

Neuroprotection with metformin and thymoquinone against ethanol-induced apoptotic neurodegeneration in prenatal rat cortical neurons

Ikram Ullah¹, Najeeb Ullah^{1,2}, Muhammad Imran Naseer^{1,3}, Hae Young Lee¹ and Myeong OK Kim^{1*}

Abstract

Background: Exposure to ethanol during early development triggers severe neuronal death by activating multiple stress pathways and causes neurological disorders, such as fetal alcohol effects or fetal alcohol syndrome. This study investigated the effect of ethanol on intracellular events that predispose developing neurons for apoptosis via calcium-mediated signaling. Although the underlying molecular mechanisms of ethanol neurotoxicity are not completely determined, mitochondrial dysfunction, altered calcium homeostasis and apoptosis-related proteins have been implicated in ethanol neurotoxicity. The present study was designed to evaluate the neuroprotective mechanisms of metformin (Met) and thymoquinone (TQ) during ethanol toxicity in rat prenatal cortical neurons at gestational day (GD) 17.5.

Results: We found that Met and TQ, separately and synergistically, increased cell viability after ethanol (100 mM) exposure for 12 hours and attenuated the elevation of cytosolic free calcium $[Ca^{2+}]_c$. Furthermore, Met and TQ maintained normal physiological mitochondrial transmembrane potential ($\Delta\Psi_M$), which is typically lowered by ethanol exposure. Increased cytosolic free $[Ca^{2+}]_c$ and lowered mitochondrial transmembrane potential after ethanol exposure significantly decreased the expression of a key anti-apoptotic protein (Bcl-2), increased expression of Bax, and stimulated the release of cytochrome-c from mitochondria. Met and TQ treatment inhibited the apoptotic cascade by increasing Bcl-2 expression. These compounds also repressed the activation of caspase-9 and caspase-3 and reduced the cleavage of PARP-1. Morphological conformation of cell death was assessed by TUNEL, Fluoro-Jade-B, and PI staining. These staining methods demonstrated more cell death after ethanol treatment, while Met, TQ or Met plus TQ prevented ethanol-induced apoptotic cell death.

Conclusion: These findings suggested that Met and TQ are strong protective agents against ethanol-induced neuronal apoptosis in primary rat cortical neurons. The collective data demonstrated that Met and TQ have the potential to ameliorate ethanol neurotoxicity and revealed a possible protective target mechanism for the damaging effects of ethanol during early brain development.

Background

Exposure to ethanol during brain development might provoke neurodevelopmental defects that are referred to as fetal alcohol effects (FAE) or fetal alcohol syndrome (FAS) [1]. Ethanol potentially damages the developing brain by affecting neurogenesis, cell migration, or cell

survival via different intracellular signaling pathways in prenatal rat cortical and hippocampal neurons [2-4]. Neurodegeneration presents with reduced brain mass and neurobehavioral disturbances in many neurological disorders, and cell death processes are associated with the activation of caspase-3, an executioner protease that is active during apoptotic cell death [5,6]. Apoptosis is a normal process in the developing brain; for optimal development, more than 50% of the original neurons must undergo programmed cell death or apoptosis [7]. Apoptosis is an indispensable mechanism for cellular

* Correspondence: mokim@gsnu.ac.kr

¹Department of Biology, College of Natural Sciences (RINS) and Applied Life Science (BK 21), Gyeongsang National University, Jinju, 660-701, Republic of Korea

Full list of author information is available at the end of the article

migration and differentiation during the formation of the developing central nervous system. In particular, Bax and Bcl-2 are major apoptotic proteins and act as an apoptotic inducer and inhibitor, respectively [8,9].

Mitochondria play an important role in apoptosis under a variety of pro-apoptotic conditions, such as oxidative stress and cytochrome-c release. The latter is a key event in the activation of caspase-3, a pivotal downstream step in apoptosis initiation [10,11]. Research has demonstrated protective roles for vitamin E and brain derived-neurotrophic factor (BDNF) against ethanol-induced apoptosis, which causes the generation of reactive oxygen species (ROS) and disrupts mitochondrial membrane potential ($\Delta\psi_M$) [12]. Cellular or sustained Ca^{2+} overload is also an important mechanism that contributes to ethanol-induced programmed cell death through the activation of post-mitochondrial events involving caspase activation [13,14].

Different therapeutic approaches have been investigated to overcome neurodegenerative diseases [15]. Some approaches consist of drugs that act directly on mitochondrial function, such as cyclosporine A, which is a permeability transition pore (PTP) reference blocker [16]. It was reported that the antidiabetic drug metformin (Met) prevents PTP opening and subsequent cell death in various endothelial cell types exposed to high glucose levels [17]. Recently, it was also reported that Met can interrupt the apoptotic cascade in a model of etoposide-induced cell death by inhibiting PTP opening, blocking the release of cytochrome-c and preventing subsequent cell death [18]. This finding is of great clinical interest because Met, beyond its anti-hyperglycemic function, can also act as a pharmacological agent in the treatment of neurodegenerative disorders. In this study, we showed that Met is substantially neuroprotective at a therapeutic dose against ethanol-induced cell death in rat primary cortical neurons.

Thymoquinone (TQ), the active component of *Nigella sativa* (NS) seeds, has broad and versatile pharmacological effects. These effects include strong antioxidant activity against free radical-generating agents, such as doxorubicin-induced cardiotoxicity [19,20]. Recently, it was reported that NS extract and TQ protect against cell death induced by serum/glucose deprivation in PC12 cells via a direct reduction in intracellular ROS [21] and in chronic toluene-induced neurodegeneration in the hippocampus [22]. Recent studies reported that TQ prevented brain damage in a model of transient forebrain ischemia in the rat hippocampus. TQ stimulated resistance to oxidative stress by decreasing the elevated levels of malondialdehyde, glutathione (GSH) contents, catalase and superoxide dismutase (SOD) [23]. A protective role of TQ was reported in 1-methyl-4-phenylpyridinium (MPP)-treated primary dopaminergic

cultures and a primary Parkinson's disease model involving rotenone and neuroinflammatory mechanisms [24].

Previously, we reported that antioxidant vitamin C was neuroprotective against ethanol and nicotine via the modulation of GABA_B receptor and protein kinase A (PKA- α) expression in prenatal and postnatal rat brain [25,26]. Thus, this study was designed to investigate the molecular mechanisms involved in TQ and Met neuroprotection against ethanol-induced apoptosis. We investigated how Met and TQ regulate Ca^{2+} dysregulation, mitochondrial dysfunction, cytochrome-c release, caspase activation and the Bcl-2 family of proteins during ethanol-induced neuronal apoptosis.

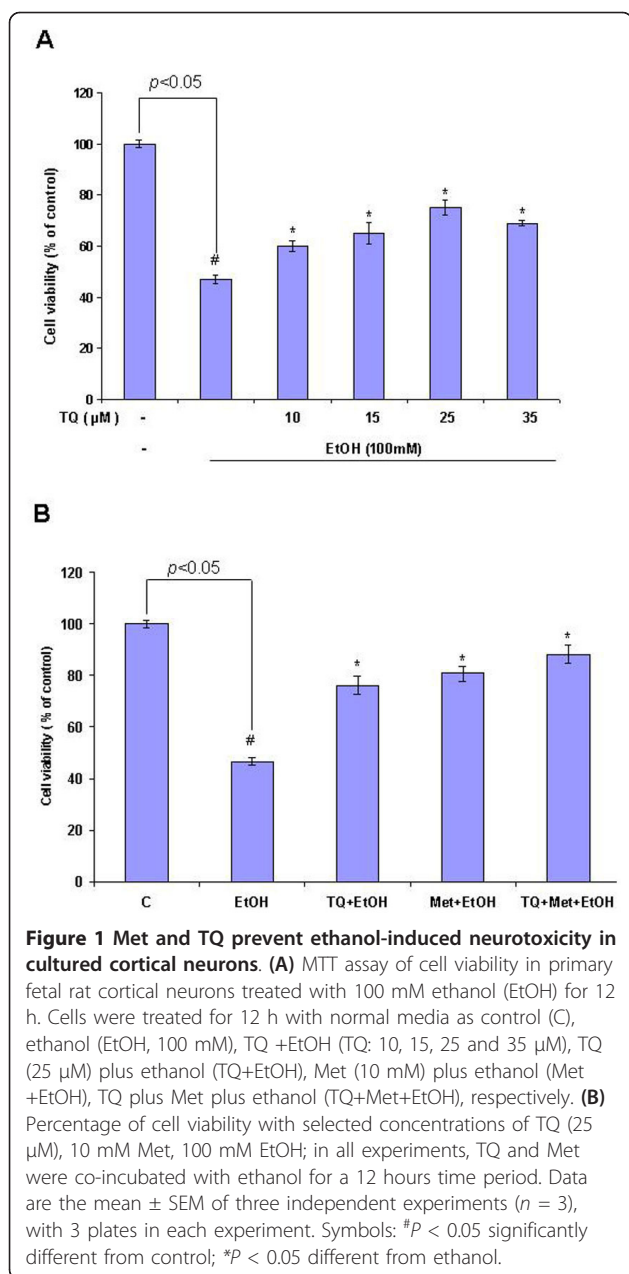
Results

Effect of TQ and Met on ethanol-induced cell death in primary rat cortical neurons

Cortical neurons were treated with ethanol (100 mM), TQ (10, 15, 25 and 35 μ M) and Met (10 mM) in different combinations for 12 h, and cell viability was measured by 3-[4,5-dimethylthiazol-2]-2,5 diphenyltetrazolium bromide (MTT) assay. TQ and Met were co-incubated with ethanol in all experimental groups for 12 h. Cortical neurons exposed to ethanol exhibited a decrease in cell viability after 12 h incubation, but cotreatment with TQ at different concentrations reversed cell loss. All of the TQ concentrations tested were effective, but 25 μ M was the most efficacious (Figure 1A). In the context of ethanol treatment, cells supplemented with TQ- or Met-only showed increased cell viability, while TQ coadministered with Met had an even stronger effect on cell viability, which indicated a synergistic effect (Figure 1B).

TQ and Met prevent ethanol-induced dysregulation of intracellular free $[Ca^{2+}]$ homeostasis

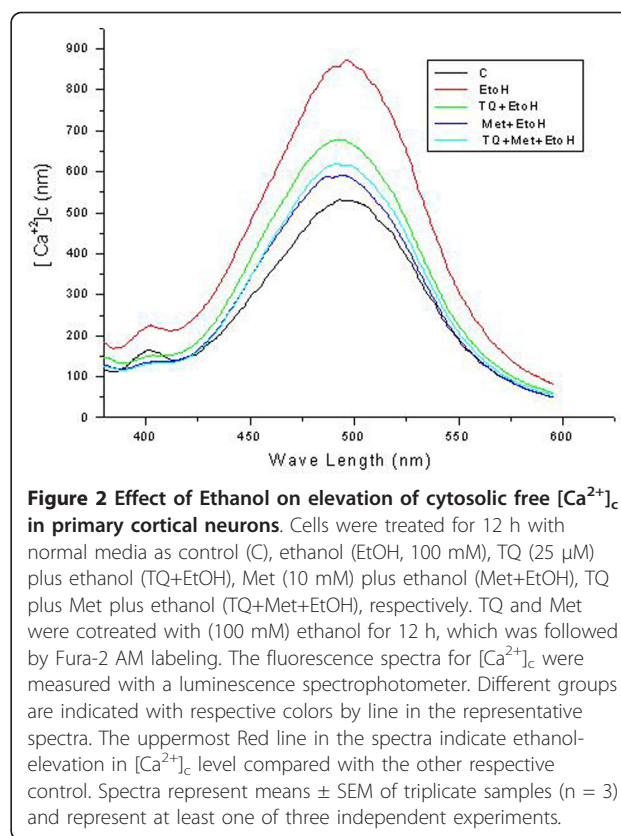
One potential mechanism of ethanol-induced neurotoxicity is through the dysregulation of cytoplasmic Ca^{2+} concentration. Cytoplasmic Ca^{2+} concentration governs neurotransmitter release and neural development, while its dysregulation results in neuronal cell death [4]. Abnormalities in Ca^{2+} homeostasis, particularly overload, was linked to neural apoptosis in the present study. Cytosolic free Ca^{2+} was measured with a fluorescent Ca^{2+} indicator, Fura-2, after a 12 h drug treatment. Ethanol treatment increased cytosolic free Ca^{2+} with peak levels at 869 ± 16.37 nM compared with control having peak level 531 ± 8.06 nM. TQ and Met cotreatment with ethanol restored normal Ca^{2+} levels with a peak levels 677 ± 6.19 nM and 590 ± 5.54 nM respectively. TQ and Met in combination also prevented ethanol-induced Ca^{2+} elevation with a peak level 617 ± 4.32 nM (Figure 2). We observed that the intracellular Ca^{2+} concentration was significantly elevated in cells exposed to ethanol (100 mM) for 12 h compared to control,



which led to neuronal cell death. TQ and Met reversed this effect that ethanol had on intracellular Ca^{2+} , which suggested that they have a protective role in maintaining Ca^{2+} homeostasis.

TQ and Met maintain mitochondrial transmembrane potential ($\Delta\psi_M$) and prevent neuronal cell death

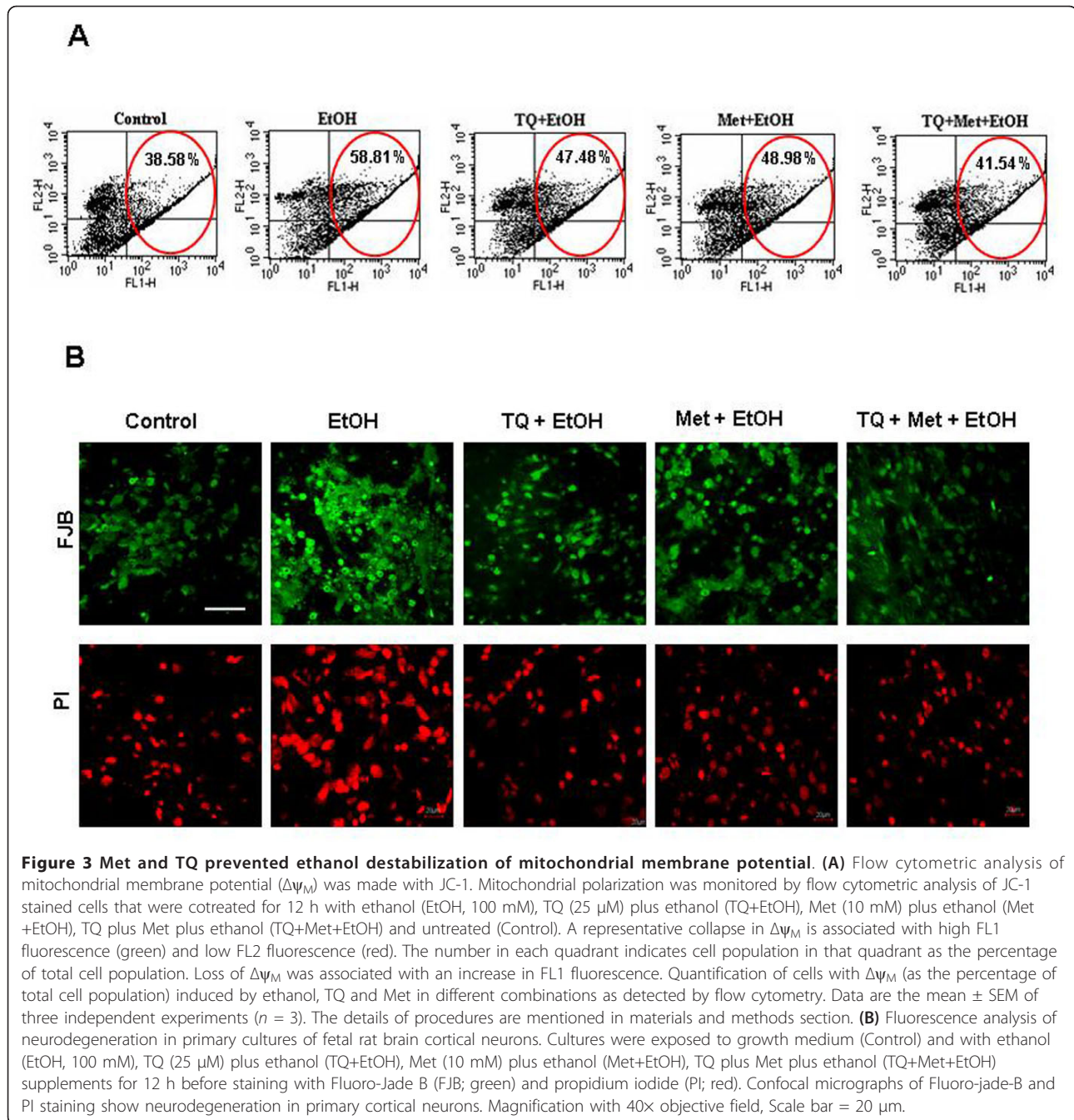
We examined mitochondrial membrane potential ($\Delta\psi_M$) using flow cytometry that measured the intensity of green and red fluorescence. We used a computer-based program that calculated the ratio of green to red fluorescence after 12 h of drug treatment. Ethanol (EtOH)



treatment resulted in a higher ratio of green/red FL1 fluorescence $58.8 \pm 1.96\%$ ($P < 0.05$) relative to control $38.5 \pm 1.81\%$. Lower FL1 fluorescence, relative to ethanol-alone group, for TQ+EtOH $47.4 \pm 1.55\%$ ($P < 0.05$), Met+ EtOH $48.9 \pm 2.31\%$ ($P < 0.05$) or TQ+Met+EtOH $41.5 \pm 1.14\%$ ($P < 0.05$) indicated that Met and TQ block apoptosis by preventing $\Delta\psi_M$ disruption (Figure 3A). TQ and Met resulted in the appearance of a remarkable subpopulation of cells with normal $\Delta\psi_M$, which further supported the protective effect of TQ and Met against ethanol. Fluoro-Jade-B (FJB) and propidium iodide (PI) staining was performed to assess morphological cell death induced by ethanol. After 12 h, the ethanol treatment group had a greater number of dead cells; FJB and PI staining was markedly condensed with higher intensity compared to control. Cotreatment with TQ and Met reversed the effects of ethanol (Figure 3B).

Effect of TQ and Met on ethanol-induced changes in Bcl-2 and Bax expression in cortical neurons

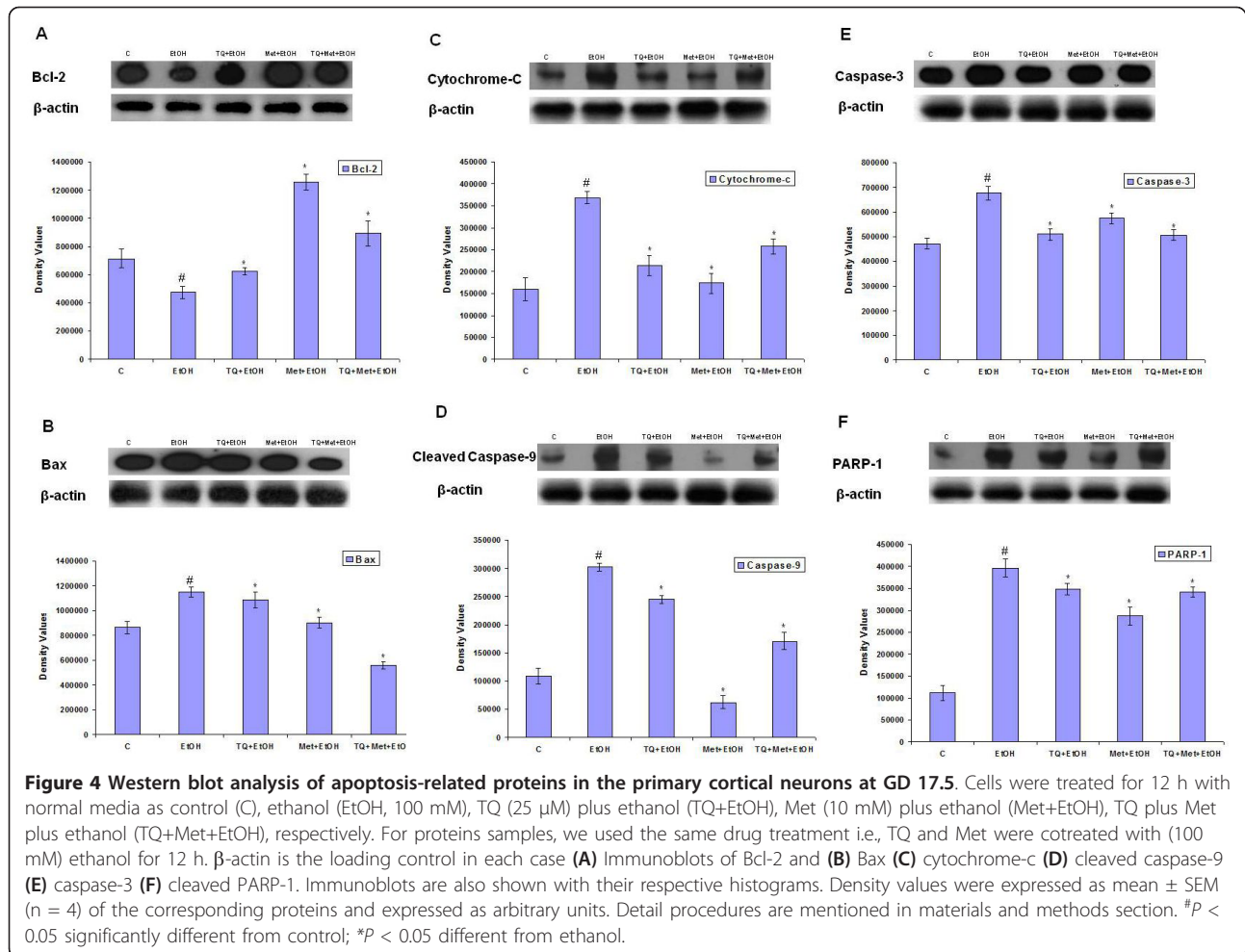
Bcl-2 and Bax are members of a family of cytoplasmic proteins that regulate apoptosis. To observe the protective effect of TQ and Met against ethanol-induced apoptosis, we assayed Bcl-2 and Bax expression levels by western blot analysis. Western blot analysis demonstrated that the expression of anti-apoptotic Bcl-2 was



enhanced by TQ and Met treatment but reduced by ethanol (Figure 4A). In contrast, the pro-apoptotic Bax was upregulated by ethanol but downregulated when cotreated with TQ, Met or TQ plus Met (Figure 4B). Our results showed that TQ and Met prevented neuronal apoptosis by decreasing the expression of pro-apoptotic Bax protein while increasing the expression of anti-apoptotic Bcl-2 protein in primary cortical neurons.

TQ and Met reduced the release of cytochrome-c into the cytosol and inhibited caspase-9 activation induced by ethanol

The mitochondrial apoptotic cascade requires the release of cytochrome-c, an intermitochondrial membrane protein, into the cytosol. This release activates caspases-9 and caspase-3, which causes neuronal cell death [27]. Our results revealed that ethanol exposure increased the expression cytosolic cytochrome-c levels



relative to control. Interestingly, TQ and Met supplementation reversed this trend. Cells treated with ethanol along with TQ or Met had significantly reduced cytochrome-c expression levels relative to the ethanol treatment group (Figure 4C). Caspases are critical mediators of apoptosis in mammalian cells [28]. We analyzed the expression of cleaved caspase-9 after ethanol exposure. Our results showed that ethanol significantly increased cleaved caspase-9 in primary cortical neurons relative to the control group, while cotreatment of ethanol with TQ or Met resulted in significantly lower cleaved caspase-9 levels compared to the ethanol treatment group (Figure 4D).

Effect of TQ and Met on ethanol-induced expression of caspase-3 and cleavage of PARP-1 in primary rat cortical neurons

Caspase-3 is one of the key executioners of apoptosis and is responsible, either partially or completely, for the proteolytic cleavage of many key proteins [28]. Furthermore, the involvement of caspase-3 in cell death was

analyzed by western blotting. Ethanol induced the activation of caspase-3, while TQ and Met effectively decreased ethanol-induced expression of caspase-3 in primary cortical neurons (Figure 4E). Furthermore, we determined whether the elevated levels of activated caspase-3 that were observed in the cortical neurons treated with ethanol led to the cleavage of PARP-1, which occurs during apoptosis. Despite its function in DNA repair, overactivation of PARP-1 during neuronal excitotoxicity induces cell death [29]. The level of cleaved PARP-1 in ethanol-treated cortical neurons was significantly enhanced compared to control neurons. In the setting of ethanol, TQ and Met treatment resulted in a remarkable decrease in the level of PARP-1 and decreased its 89 kDa cleaved products in the cortical neurons (Figure 4F). Our results collectively demonstrated that an overall increase in PARP-1 cleavage occurred in the cortical neurons after exposure to ethanol. TQ and Met significantly reduced the levels of ethanol-induced cleaved PARP-1 relative to the control groups (Figure 4F).

Effect of TQ and Met treatment on ethanol-induced neurodegeneration in the primary cultured cortical neurons after 12 h

DNA damage is one of the main features of apoptosis. Visualization of DNA damage is accomplished with TUNEL staining, which is an assay for DNA breaks based on enzymatic labeling of free 3' DNA ends. The nuclear morphology of apoptotic neuronal cells was evaluated with TUNEL counterstained with DAPI (TUNEL: DNA fragmentation, DAPI: nuclear marker). Ethanol exposure increased the number of TUNEL-positive neurons by (58%) compared with control. TQ with ethanol showed significantly reduced the TUNEL positive cells by (38%) compared to the ethanol treatment group. Further Met with ethanol reduced the percentage of apoptotic cells by (37%) and TQ plus Met with ethanol (52%) compared to the cells treated with ethanol. Although there are DAPI-positive cells in control and ethanol-treated group, the pattern of staining differs between the groups. Ethanol-treated cells exhibited shrunken nuclei and abnormal cell morphology (Figure 5).

Discussion

Ethanol exposure during prenatal development causes a wide range of structural and functional brain abnormalities [30,31]. It has already been established that ethanol depletes many cell populations during development, but not all populations of neurons in the brain are susceptible; furthermore, ethanol increases neuronal apoptosis, which causes malformations, including the loss of brain mass [1,32,33]. Ethanol increases ROS generation, which can trigger apoptotic cell death pathways through the activation of caspases [34-36]. The present study provides strong evidence for a direct neuroprotective role of Met and TQ. The current investigation indicated that cotreatment of ethanol with Met, TQ or Met plus TQ protects cortical neurons from the apoptosis triggered by ethanol during early development.

The evidence regarding neuroprotection with the anti-diabetic drug Met is supported by studies that state its beneficial effects in various models of toxicity. It was reported that Met can protect against neurotoxicity, in addition to its antidiabetic effect, and it has a number of important, diverse functions, including in polycystic ovarian syndrome, induction of osteoblasts via AMP-activated protein kinase (AMPK) signaling pathway along with subsequent enhancement of nitric oxide synthase (eNOS) and bone morphogenetic-2 protein (BMP-2) production [37,38]. Met modulates AMPK and mitogen-activated protein kinase (MAPK) pathways to induce bone formation, which mimics the intracellular metabolic changes associated with nutritional deprivation [39,40]. An additional target of Met is the respiratory chain complex I of the mitochondria, which

activates AMPK via changes in mitochondrial reactive nitrogen species [41,42]. However, we provide here a new and potential mechanism that supports a protective role of Met on apoptosis that is likely due to it maintaining mitochondria integrity and reducing $[Ca^{2+}]_c$ overload as well as subsequent cell death [18].

Furthermore, we studied TQ, which is a natural compound with different beneficial pharmacologic effects against various cytotoxic insults. These beneficial effects have been observed *in vitro* and *in vivo* with degeneration caused by L-dopa in SH-SY5Y human neuroblastoma cells, serum/glucose deprivation induced cell death (SGD) and chronic toluene exposure [21,22,43]. In general, previously published reports showed that TQ mainly functions through its antioxidant mechanism, and it has been used as a protective agent in multiple toxicity models, such as doxorubicin-induced cardiotoxicity, cisplatin-induced nephrotoxicity and ischemic neuronal cell death [21,23,44].

We found that administration of Met and TQ in selected doses has the capability to maintain normal cell viability against ethanol. TQ, Met or a combination of both reduced the cell death induced by ethanol (Figure 1A, B). Calcium homeostasis is an important for normal cellular physiological function, and even a minute variation in normal levels can provoke abnormal cellular function and cell death [45]. This $[Ca^{2+}]_c$ overload further disrupts mitochondrial membrane potential ($\Delta\psi_M$), which leads to neuronal apoptosis. In this study, we observed a two-fold increase in $[Ca^{2+}]_c$ levels upon ethanol treatment compared to control cells. This cellular accumulation of calcium possibly induced cell death. Previously, a similar increase in intracellular $[Ca^{2+}]_c$ was observed after a short (20 min) exposure to ethanol in prenatal primary neuronal cells [4], fetal hypothalamic cells and cervical ganglion neurons [45,46]. TQ and Met reduced elevated $[Ca^{2+}]_c$ levels (Figure 2). TQ and Met modulate and stabilize $\Delta\psi_M$. Our findings that showed ethanol-induced alterations in mitochondrial function were also in agreement with a recent study that demonstrated that vitamin E and BDNF have neuroprotective and modulating roles during ethanol toxicity in cerebellar granule cells. The mechanism of neuroprotection was similar to Met and TQ involved the stabilization of mitochondrial membrane potential and inhibition of apoptotic cascade [12].

The Bcl-2 protein family plays an important role in apoptotic signal transduction by regulating mitochondrial function [47]. It was reported that mitochondrial cytochrome-c is released into the cytosol through the permeability transition pore, which is regulated by Bcl-2 family proteins [47]. The pro-apoptotic protein Bax destabilizes $\Delta\psi_M$ and facilitates cytochrome-c release [48], while the anti-apoptotic protein Bcl-2 functions to

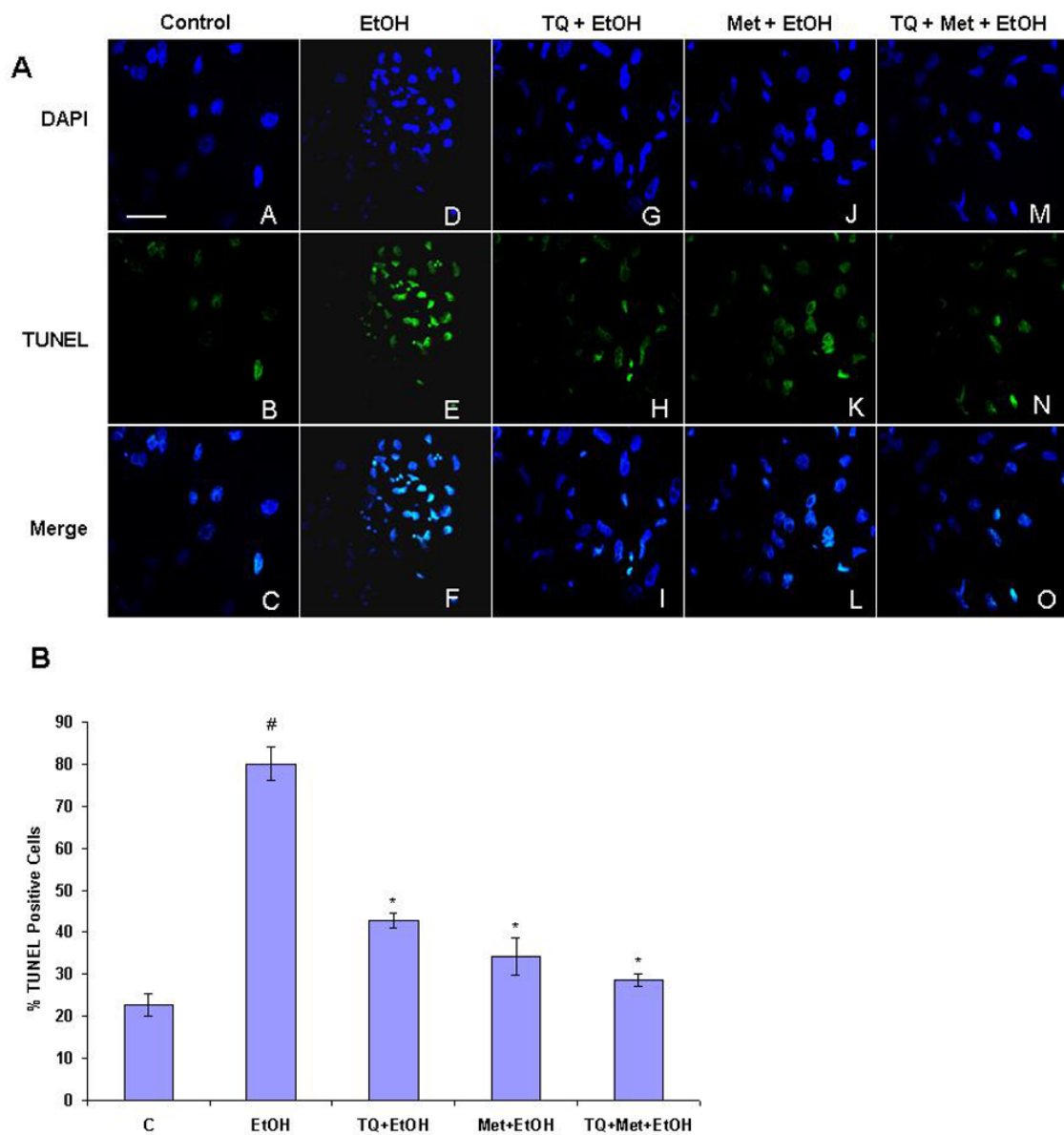


Figure 5 Morphological assessment of ethanol-induced apoptosis via TUNEL assay. (A) Cells were treated for 12 h with normal media as control (C), ethanol (EtOH, 100 mM), TQ (25 μ M) plus ethanol (TQ+EtOH), Met (10 mM) plus ethanol (Met+EtOH), TQ plus Met plus ethanol (TQ +Met+EtOH), respectively. Effects of ethanol, TQ plus ethanol, Met plus ethanol and TQ plus Met plus ethanol on apoptotic death in prenatal rat cortical neurons was visualized with TUNEL and DAPI stains. Representative photomicrographs of TUNEL staining show apoptotic neurons after ethanol administration followed by TQ and Met. TQ and Met treatment effectively blocked ethanol-induced apoptosis, as evidenced from the lack of TUNEL-positive cells. Panels A-O display TUNEL-stained cells observed by confocal microscopy at higher magnifications with a 40 \times objective field, Scale bar = 20 μ m. **(B):** The percentage of TUNEL positive cells in each case were counted and the cumulative data from three independent experiments is shown here as mean \pm SEM (n = 3). [#]*P* < 0.05 significantly different from control; ^{*}*P* < 0.05 different from ethanol.

restore membrane potential and block cytochrome-c release. In our findings, we found that TQ and Met increased Bcl-2 levels in primary cortical neurons exposed to ethanol. TUNEL and western blot analysis demonstrated that TQ and Met cotreatment protects neurons by lowering the expression of caspase-3, cytochrome-c, cleaved caspase-9 while also increasing the

expression of Bcl-2 compared to cells treated with ethanol alone. This finding implied that TQ and Met potentially prevents apoptosis by regulating the mitochondrial pathway.

PARP-1 functions in apoptosis by acting as a survival factor and a death promoter, depending on the duration and severity of DNA damage [49]. We observed cleavage

of PARP-1 to its respective 89 kDa fragment after a 12 h exposure to ethanol. In Met and TQ treatment groups, fragmentation of PARP-1 was significantly reduced. In support of our results, other investigators noted caspase-dependent cleavage of PARP-1 that was induced by an ethanol concentration (4.0 mg/ml), similar to the one used in this study, for 0, 4, 12 and 24 h time periods [50]. The potential benefit of Met and TQ on primary neuronal cell viability was evaluated by TUNEL assay. Met and TQ decreased DNA damage and reduced the number of TUNEL-positive cells produced by ethanol treatment. A study using an analogous experimental approach demonstrated similar DNA damage induced by ethanol in cultured cortical neurons [50].

The present findings supported a conclusion that ethanol disrupts $\Delta\Psi_M$, sustains Ca^{2+} overload and triggers an apoptotic cascade. Met and TQ inhibition of ethanol-induced cell death in primary rat cortical neurons might occur via an antioxidant mechanism that maintains mitochondrial integrity. Since Met and TQ are safe and non-toxic, more studies should be conducted to explain their synergistic actions with other drugs.

Conclusion

In summary, this study demonstrated that TQ and Met exert a neuroprotective effect by decreasing ethanol-mediated mitochondria-dependent apoptosis. Although additional studies are needed to determine the molecular mechanisms underlying TQ and Met actions, TQ and Met are readily available and safe agents, so they could eventually be used as neuroprotectants against ethanol-induced neurodegeneration during early development.

Methods

Animal treatment

Female ($n = 12$) Sprague-Dawley rats (250 g, Gyeongsang National University, Neurobiology Laboratory, Chinju, South Korea) were housed in a temperature-controlled environment (light cycle: 06:00-20:00 h) with food ad libitum. Time of pregnancy was assigned from the day of insemination equal to gestational day (GD) 0.5. After GD 17.5 pregnant Sprague-Dawley rats were killed by decapitation and after an i.v. injection of pentobarbital sodium (3 mg/100 g body weight). Animals were treated in accordance with standard guidelines for laboratory animal care. Experimental procedures were approved by the local animal ethics committee of the Division of Applied Life Sciences, Department of Biology, Gyeongsang National University, South Korea.

Primary cell culture and drug treatment

Cultures were prepared from the cortices of prenatal rats at GD 17.5. Pooled cortical brain tissue was treated

with 0.25% trypsin-EDTA for 20 min and dissociated by mechanical trituration in ice-cold calcium-free and magnesium-free Hank's balanced salt solution (pH 7.4). After pelleting by centrifugation, cells were plated (1×10^6 cells/ml) in chamber slides or cell culture plates that were pre-coated with polylysine (0.02 g/l). Culture medium consisted of Dulbecco's modified Eagle medium (DMEM) with 10% heat-inactivated fetal bovine serum, 1 mM pyruvate, 4.2 mM sodium bicarbonate, 20 mM HEPES, 0.3 g/L bovine serum albumin, 50 U/ml penicillin, and 50 mg/L streptomycin. Cultures were maintained at 37°C in a humidified atmosphere of 5% CO₂ and 95% air. Neuroglia cells were inhibited with media containing 100 μ M Cytosine β -D-Arabino Furanoside (Sigma) for 12 h. After 4 days, cortical neurons were assigned to of five groups: 1) Control (C): cells incubated with normal DMEM medium; 2) Ethanol treatment alone (EtOH): treated with media containing 100 mM ethanol; 3) TQ plus EtOH cotreatment (concurrently); 4) Met plus EtOH cotreatment (concurrently); 5) TQ plus Met plus EtOH cotreatment (concurrently). The TQ dose (25 μ M) was selected on the basis of MTT assay, and the Met dose (10 mM) was from unpublished lab data; these were used in different groups and combinations. All drug treatment groups were incubated for 12 h *in vitro* selected on the basis of preliminary experiments carried out and on the basis of that we selected 12 h as an optimum condition to carry out the final experiments reported here. In all experiments 100 mM ethanol concentration was used selected from of our previously published studies [4].

Cell viability

The logarithmic growth phase of primary neuronal cells was taken for growth assays using 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide (MTT). Primary cortical neurons were seeded onto 96-well plates (1×10^5 cells/well) in 200 μ l of DMEM media (control). DMEM media with ethanol (100 mM), TQ (10, 15, 25 and 35 μ M) and Met (10 mM) in different combinations were incubated at 37°C for 12 h in a humidified 5% CO₂ incubator. TQ and Met were cotreated with ethanol in all experimental groups. After incubation, cell viability was determined by adding MTT (5 mg/ml in phosphate buffer saline, PBS) to each well and incubating these cells for 4 h at 37°C. Formazan dissolved in organic solvent was added, to the cells. Plates were placed on a shaker and agitated for 10 to 20 min. Plates were read at 550 to 570 nm (L1) and 620 to 650 nm (L2) on scanning microplate reader spectrophotometer; 620 to 650 nm absorbance results measured cell debris and well imperfections. The final optical density (OD) obtained (OD = L1-L2) was used to calculate the percentage of cell survival, which was expressed as absorbance treated wells/absorbance control wells \times 100%.

Intracellular free Ca²⁺ measurement

The intracellular Ca²⁺ concentration was measured with the fluorescent Ca²⁺ indicator Fura-2 acetoxymethyl ester [51]. After four days of growth, cells were treated with normal media (control), ethanol (100 mM), TQ (25 μM) plus ethanol, Met (10 mM) plus ethanol, or TQ plus Met in combination with ethanol for 12 h. After treatment, cultured cortical neurons (triplicate plates; each containing 1 × 10⁶ cells) were washed twice with Krebs buffer and then incubated in DMEM media containing 5 μM Fura-2 AM at 37°C in a humidified incubator with 5% CO₂ for 60 min. Cells were washed twice with Locke's solution (pH 7.8), and Fura-2 fluorescence signals of [Ca²⁺]_c were measured with a luminescence spectrophotometer (LS50B, Perkin Elmer, Boston, MA). Excitation light from a Xenon lamp was alternated between 340 and 380 nm band-pass filters and the fluorescence emitted at 510 nm was revealed by a photon-counting photomultiplier. The 340 nm/380 nm fluorescence ratio, averaged over a period of 2 s, was measured. Fluorescence signals were acquired, stored and analyzed using a computer with universal imaging software or a MicroVax II computer with origin 7 software. Intracellular calcium was determined using the ratio method as mentioned above and from the Grynkiewicz equation [52].

$$[Ca] = K_d \times \frac{(R - R_{min})}{(R_{max} - R)} \times \frac{Sf2}{Sb2}$$

K_d: dissociation constant of the Fura-2 Ca²⁺ interaction to be 225 nM in the cytosolic environment; R: fluorescence ratio at 340 and 380 nm; R_{min}: ratio with zero Ca²⁺; R_{max}: ratio with saturating Ca²⁺ (using calcium chloride); Sf2: fluorescence at 380 nm with zero Ca²⁺; Sb2: fluorescence at 380 nm with saturating Ca²⁺.

Measurement of mitochondrial membrane potential with JC-1

Mitochondrial membrane potential (Δψ_M) was monitored using JC-1 Δψ_M detection kit (Biotium Inc., Hayward, CA, USA), according to the manufacturer's protocol. JC-1 emits either green or red fluorescence, depending on the Δψ_M. A green signal indicated mitochondria depolarization, while a red signal indicated polarized mitochondria [53]. Thus, the shift from red to green fluorescence was considered a reliable indicator of a drop in Δψ_M. After 4 days, neuronal cells were treated with normal media (control; C), ethanol (100 mM; EtOH), ethanol plus TQ (25 μM), Met plus ethanol and TQ plus Met plus ethanol in different groups and combinations. All drug treatment groups were incubated for 12 h *in vitro* at the same time at 37°C. After drug treatment, cells in triplicate culture were washed with PBS

twice, harvested and stained with JC-1 reagents at 37°C for 15 min. Cells were resuspended twice in 1× assay buffer. JC-1 aggregates in healthy mitochondria emit red fluorescence at 590 nm. JC-1 monomers that leaked from stressed mitochondria emit green fluorescence at 530 nm. Red and green fluorescence were measured in the green (FL-1) and red (FL-2) channels of a flow cytometer. Changes in Δψ_M were measured at the single-cell level with a FACS Caliber flow cytometer (Becton Dickinson, San Jose, CA, USA). In total, 10,000 cells were acquired for analysis by Cell Quest software, version 3.0 (Becton Dickinson, San Jose, CA, USA), and quantification of cells with low Δψ_M, as the percentage of total cell population, was performed.

Western blotting

Western blot analysis was done as previously described with some modifications [54]. Immunoreactions were performed with primary antibodies: Bax and Bcl-2 (Rabbit polyclonal, Santa Cruz Biotechnology; 1:1000, 24 h, 4°C), cytochrome-c (Goat, Santa Cruz Biotechnology; 1:1000, 24 h, 4°C), PARP-1 (Mouse monoclonal, Santa Cruz Biotechnology; 1:1000, 24 h, 4°C), caspase-3 and caspase-9 antibody (Rabbit, Cell signaling 1:1000 24 h, 4°C). After rinsing the blots, horseradish peroxidase-conjugated goat anti-mouse, mouse anti-goat or goat anti-rabbit IgG-HRPs (Santa Cruz Biotech 1:1000) were incubated with the blots for 2 h at room temperature. Immunoreactions were also performed using β-actin antibody (Cell Signaling) as loading controls. Proteins were detected by chemiluminescence using an ECL-detecting reagent (Amersham Pharmacia Biotech, western blotting detection reagents) with the manufacturer's protocol. Chemiluminescent blots were then exposed to X-ray film. Exposed X-ray film was scanned, and optical densities were analyzed by densitometry using the computer-based SigmaGel, version 1.0 (Jandel Scientific, San Rafael, Chicago, USA).

TUNEL and DAPI staining

To assay apoptosis, cells were stained with TUNEL (GenScript Corporation, USA) and counterstained using 4',6-diamidino-2-phenylindole (DAPI). In situ detection of apoptotic cell death was performed using terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end-labeling (TUNEL) in cortical neuron cultures as previously done with a little modification [13]. TUNEL staining was performed, according to the supplier's recommendations, using the In Situ Cell Death Detection kit for Fluorescein (GeneScript, USA). Cells were also incubated with DAPI (Molecular Probes, Eugene, OR, USA) for 10 min at room temperature and then rinsed with distilled water. Glass cover slips were mounted on glass slides with mounting medium.

A DAPI filter was used to detect DAPI staining (blue color), and an FITC filter was used to detect TUNEL staining (green color). TUNEL-positive (green) and DAPI-positive (blue) staining patterns were acquired with a confocal laser scanning microscope (Fluoview FV 1000, Olympus, Japan). TUNEL-positive cells in the different regions of each slide were counted by an observer who was blinded to the treatment conditions.

Fluoro-Jade-B (FJB) and PI staining

Fluoro-Jade-B (FJB) staining was performed as previously described [55]. Immunofluorescence was performed on GD 17.5 primary cortical neuron cultures grown *in vitro* on poly-D-lysine-coated chamber slides. Four-day-old cultures were treated for 12 h at 37°C as indicated in the figure legends. Treated cultures were fixed for 5 min with 4% paraformaldehyde in PBS and stored at -70°C. Slides were air dried for 3 h and then subjected to the following treatments in order: 10 min in 0.06% potassium permanganate solution, distilled water rinse, 20 min in 0.1% acetic acid containing 0.0004% FJB (Calbiochem, San Diego, California, USA) and three washes in distilled water. The slides were allowed to dry at 55°C for 10 min and viewed under an FITC filter in a confocal microscope (Olympus Fluoview FV1000, Japan). For propidium iodide (PI) staining, slides were dipped in PI solution (1 µg/ml) in PBS with gentle mixing for 20 min at room temperature and washed twice with PBS for 10 min. Glass cover slips were mounted on the slides with mounting medium.

Data analysis and statistics

The objective bands on western blots were scanned and analyzed by densitometry using a computer with the SigmaGel System (SPSS Inc., Chicago, IL). Density values were expressed as mean ± SEM. One-way ANOVA followed by Tukey-Kramer multiple comparisons test was performed to determine the significance of differences between relevant treatment groups. In every case, the acceptance level for statistical significance was $P < 0.05$.

Abbreviations

Met: Metformin; TQ: Thymoquinone; FAS: fetal alcohol syndrome; PTZ: pentylenetetrazol; GD: gestational days; DEPC: diethyl pyrocarbonate; NBP: 4% neutral buffer paraformaldehyde; DMEM: Dulbecco's modified Eagle medium; MTT: 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide; PCD: programmed cell death; PTP: permeability transition pore; NS: *Nigella sativa*.

Acknowledgements

This work was supported by a grant from the Next Generation Biogreen 21 Program (PJ008075 and Agenda PJ007361) of Rural Development Administration and NRF (2010-0000206 and 2009-353-C00062) funded by the Korean government.

Author details

¹Department of Biology, College of Natural Sciences (RINS) and Applied Life Science (BK 21), Gyeongsang National University, Jinju, 660-701, Republic of Korea. ²Institute of Basic Medical Sciences, Khyber Medical University, Peshawar, Khyber Pakhtoonkhwa, Pakistan. ³Center of Excellence in Genomic Medicine (CEGMR), King Abdulaziz University, Jeddah, Saudi Arabia.

Authors' contributions

IKU carried out the experiments and drafted the manuscript. NAU, MIN and HYL helped in analysis and drafting the manuscript. MOK wrote discussion and conceived of the study and interpreted the data and discussion of the results. All the authors approve the final manuscript.

Received: 11 August 2011 Accepted: 19 January 2012

Published: 19 January 2012

References

1. Jones KL, Smith DW, Ulleland CN, Streissguth AP: **Pattern of malformation in offspring of chronic alcoholic mothers.** *Lancet* 1973, **1**:1267-1271.
2. Miller MW: **Effects of alcohol on the generation and migration of cerebral cortical neurons.** *Science* 1986, **233**:1308-1311.
3. Miller MW: **Mechanisms of ethanol induced neuronal death during development: from the molecule to behavior.** *Alcohol Clin Exp Res* 1996, **20**:128-132.
4. Naseer MI, Lee HY, Ullah N, Ullah I, Park MS, Kim SH, Kim MO: **Ethanol and PTZ effects on siRNA-mediated GABA_{B1} receptor: Down regulation of intracellular signaling pathway in prenatal rat cortical and hippocampal neurons.** *Synapse* 2010, **64**:181-190.
5. Carloni S, Mazzoni E, Balduini W: **Caspase-3 and calpain activities after acute and repeated ethanol administration during the rat brain growth spurt.** *J Neurochem* 2004, **89**:197-203.
6. Olney JW, Tenkova T, Dikranian K, Muglia LJ, Jermakowicz WJ: **Ethanol-induced caspase-3 activation in the in vivo developing mouse brain.** *Neurobiol Dis* 2002, **9**:205-219.
7. Sastry PS, Rao KS: **Apoptosis and the nervous system.** *J Neurochem* 2000, **74**:1-20.
8. Cory S, Adams JM: **The Bcl-2 family: Regulators of the cellular life-or-death switch.** *Nat Rev Cancer* 2002, **2**:647-656.
9. Roth KA, D'sa C: **Apoptosis and brain development.** *Ment Retard Dev Disabil Res* 2001, **7**:261-266.
10. Green DR, Reed JC: **Mitochondria and apoptosis.** *Science* 1998, **281**:1309-1312.
11. Roy S: **Caspase at the heart of the apoptotic cell death pathway.** *Chem Res Toxicol* 2000, **13**:961-962.
12. Heaton MB, Paiva M, Marsiglio KS: **Ethanol influences on Bax translocation, mitochondrial membrane potential, and reactive oxygen species generation are modulated by vitamin E and brain-derived neurotrophic factor.** *Alcohol Clin Exp Res* 2011, **6**:1-12.
13. Nilsen J, Chen S, Irwin RW, Iwamoto S, Brinton RD: **Estrogen protects neuronal cells from amyloid beta-induced apoptosis via regulation of mitochondrial proteins and function.** *BMC Neuroscience* 2006, **7**:1-14.
14. Kahraman S, Zup LS, McCarthy MM, Fiskum G: **GABAergic mechanism of profl toxicity in immature neurons.** *J Neurosurg Anesthesiol* 2008, **20**:233-240.
15. Chong ZZ, Li F, Maiese K: **Oxidative stress in the brain: Novel cellular targets that govern survival during neurodegenerative disease.** *Prog in Neurobiol* 2005, **75**:207-246.
16. Mattson MP, Kroemer G: **Mitochondria in cell death: novel targets for neuroprotection and cardioprotection.** *Trends in Mol Med* 2003, **9**:196-205.
17. Detaille D, Guigas B, Chauvin C, Batandier C, Fontaine E, Wiernsperger N, Leverve X: **Metformin prevents high glucose-induced endothelial cell death through a mitochondrial permeability transition-dependent process.** *Diabetes* 2005, **54**:2179-2187.
18. El-Mir MY, Dominique D, Gloria RV, Maria DE, Bruno G, Stephane A, Eric F, Angeles A, Xavier L: **Neuroprotective role of antidiabetic drug metformin against apoptotic cell death in primary cortical neurons.** *J Mol Neurosci* 2008, **34**:77-87.
19. Houghton PJ, Zarka R, de las Heras B, Hoult JR: **Fixed oil of nigella sativa and derived thymoquinone inhibit eicosanoid generation in leukocytes and membrane lipid peroxidation.** *Planta Med* 1995, **61**:1:33-6.

20. Nagi MN, Mansour MA: **Protective effect of thymoquinone against doxorubicin-induced cardiotoxicity in rats: a possible mechanism of protection.** *Pharmacol Res* 2000, **41**:283-289.
21. Mousavi SH, Tayarani-Najaran Z, Asghari M, Sadeghnia HR: **Protective effect of nigella sativa extract and thymoquinone on serum/glucose deprivation-induced PC12 cells death.** *Cell Mol Neurobiol* 2010, **30**:591-598.
22. Kanter M: **Nigella sativa and derived thymoquinone prevents hippocampal neurodegeneration after chronic toluene exposure in rats.** *Neurochem Res* 2008, **33**:579-588.
23. Al-Majed AA, Al-Omar FA, Nagi MN: **Neuroprotective effects of thymoquinone against transient forebrain Ischemia in the rat hippocampus.** *European Journal of Pharmacology* 2006, **543**:40-47.
24. Radad K, Moldzio R, Taha M, Rausch WD: **Thymoquinone protects dopaminergic neurons against MPP+ and rotenone.** *Phytother Res* 2009, **23**:696-700.
25. Naseer MI, Najeeb U, Ikram U, Zubair Hassan H, Yang MBC, Kim MO: **Vitamin-C protects ethanol induced apoptotic neurodegeneration in postnatal rat brain.** *Pak J Med Sci* 2009, **25**:718-722.
26. Naseer MI, Lee HY, Kim MO: **Neuroprotective effect of vitamin c against the ethanol and nicotine modulation of GABA_B receptor and PKA- α expression in prenatal rat brain.** *Synapse* 2010, **64**:467-477.
27. Li P, Nijhawan D, Budihardjo I, Srinivasula SM, Ahmad M, Alnemri ES, Wang X: **Cytochrome c and dATP-dependent formation of Apaf-1/caspase-9 complex initiates an apoptotic protease cascade.** *Cell* 1997, **91**:479-489.
28. Thornberry NA, Lazebnik Y: **Caspases: enemies within.** *Science* 1998, **281**:1312-1316.
29. Berger NA: **Poly (ADP-ribose) in the cellular response to DNA damage.** *Radiat Res* 1985, **101**:4-15.
30. Clarren SK, Smith DW: **The fetal alcohol syndrome.** *New Engl J Med* 1978, **298**:1063-1067.
31. Lindsley TA, Kerlin AM, Rising LJ: **Time-lapse analysis of ethanol's effects on axon growth in vitro.** *Dev Brain Res* 2003, **30**:191-199.
32. Miki T, Harris SJ, Wilce PA, Takeuchi Y, Bedi KS: **Effects of alcohol exposure during early life on neuron numbers in the rat hippocampus, I. Hilus neurons and granule cells.** *Hippocampus* 2001, **13**:388-398.
33. Sakata-Haga H, Sawada K, Hisano S, Fukui Y: **Administration schedule for an ethanol-containing diet in pregnancy affects types of offspring brain malformations.** *Acta Neuropathol* 2002, **104**:305-312.
34. Young C, Roth KA, Klocke BJ, West T, Holtzman DM, Labruyere J, Qin YQ, Dikranian K, Olney JW: **Role of caspase-3 in ethanol-induced developmental neurodegeneration.** *Neurobiol Dis* 2005, **20**:608-614.
35. Ramachandran V, Watts LT, Maffi SK, Chen J, Schenker S, Henderson G: **Ethanol-induced oxidative stress precedes mitochondrially mediated apoptotic death of cultured fetal cortical neurons.** *J Neurosci Res* 2003, **74**:577-588.
36. Antonio AM, Druse MJ: **Antioxidants prevent ethanol-associated apoptosis in fetal rhombencephalic neurons.** *Brain Res* 2008, **1204**:16-23.
37. Nestler JE, Jakubowicz DJ, Evans WS, Pasquali R: **Effects of metformin on spontaneous and clomiphene-induced ovulation in the polycystic ovary syndrome.** *New Engl J Med* 1998, **338**:876-80.
38. Ipppei K, Toru Y, Shozo Y, Mika Y, Toshitsugu S: **Metformin enhances the differentiation and mineralization of osteoblastic MC3T3- E1 cells via AMP kinase activation as well as eNOS and BMP-2 expression.** *Biochem and Biophys Res Commun* 2008, **375**:414-419.
39. Zhou G, Myers R, Li Y, Chen Y, et al: **Role of AMP-activated protein kinase in mechanism of metformin action.** *J Clin Invest* 2001, **108**:1105-7.
40. Shaw RJ, Lamia KA, Vasquez D, Koo SH, Bardeesy N, Depinho RA: **The kinase LKB1 mediates glucose homeostasis in liver and therapeutic effects of metformin.** *Science* 2005, **310**:1642-6.
41. Bailey CJ, Turner RC: **Metformin.** *New Engl J Med* 1998, **339**:1860-1861.
42. Zou MH, Kirkpatrick SS, Davis BJ, Nelson JS, Wiles WG, Schlattner U: **Activation of the AMP-activated protein kinase by the antidiabetic drug metformin in vivo. Role of mitochondrial reactive nitrogen species.** *J Biol Chem* 2004, **279**:43940-51.
43. Martin TM, Benghuzzi H, Tucci M: **The effect of conventional and sustained delivery of thymoquinone and levodopa on SH-SY5Y human neuroblastoma cells.** *Biomed Sci Instrum* 2006, **42**:332-337.
44. Badary OA, Nagi MN, Alshabana OA, AlShawaf HA, AlSohaibani MO, Al Bekairi AM: **Thymoquinone ameliorates the nephrotoxicity induced by cisplatin in rodents and potentiates its antitumor activity.** *Can J Physiol Pharmacol* 1997, **75**:1356-1361.
45. Simasko MS, Boyadjieva N, De A, Sarkar DK: **Effect of ethanol on calcium regulation in rat fetal hypothalamic cells in culture.** *Brain Res* 1999, **824**:89-96.
46. Xiao ZM, Li LJ, Yu SZ, Li CY, Zheng JQ: **Effects of extracellular Ca²⁺ influx and intracellular Ca²⁺ release on ethanol-induced cytoplasm Ca²⁺ overload in cultured superior cervical ganglion neurons.** *Neuroscience Letters* 2005, **390**:98-103.
47. Shy Y: **A structural view of mitochondria-mediated apoptosis.** *Nat Struct Biol* 2001, **8**:394-401.
48. Merry DE, Korsmeyer SJ: **Bcl-2 gene family in the nervous system.** *Annu Rev Neurosci* 1997, **20**:245-267.
49. Kim MY, Zhang T, Kraus WL: **Pol(ADP-ribosyl)ation by PARP-1:PAR-laying NAD in to nuclear signal.** *Genes Dev* 2005, **19**:1951-1967.
50. Cherian PP, Schenker S, Henderson G: **Ethanol-mediated DNA damage and PARP-1 apoptotic responses in cultured fetal cortical neurons.** *Alcohol Clin Exp Res* 2008, **32**:1-9.
51. Malgaroli A, Milani D, Meldolesi J, Pozzan T: **Fura-2 measurement of cytosolic free Ca²⁺ in monolayers and suspensions of various types of animal cells.** *J Cell Biol* 1987, **105**:2145-2155.
52. Grynkyewicz G, Poenie M, Tsien RY: **A new generation of Ca²⁺ indicators with greatly improved fluorescence properties.** *J Biol Chem* 1985, **260**:3440-3450.
53. Reers M, Smith TW, Chen LB: **J-aggregate formation of a carbocyanine as a quantitative fluorescence indicator of membrane potential.** *Biochemistry* 1991, **30**:4480-4486.
54. Naseer MI, Li S, Kim MO: **Maternal epileptic seizure induced by pentylenetetrazol: apoptotic neurodegeneration and decreased GABA_{B1} receptor expression in prenatal rat brain.** *Mol Brain* 2009, **2**:1-20.
55. Schmued LC, Hopkins KJ: **Fluoro-Jade B: a high affinity fluorescent marker for the localization of neuronal degeneration.** *Brain Res* 2000, **874**:123-130.

doi:10.1186/1471-2202-13-11

Cite this article as: Ullah et al.: Neuroprotection with metformin and thymoquinone against ethanol-induced apoptotic neurodegeneration in prenatal rat cortical neurons. *BMC Neuroscience* 2012 **13**:11.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

